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Effect of Miraculan (aTRIA) on some biochemical properties of tomato (*Lycopersicon esculantum* Mill.)

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Abstract

A field experiment was conducted to study the effect of miraculan on biochemical parameters in tomato (*Lycopersicon esculantum* Mill.). The different concentration of miraculan (0.5, 1.0, 2.0, 4.0, 6.0, 8.0 and 10 ppm) was applied at preflowering, flowering and post flowering stages i.e. 20, 40 and 60 DAS. The lower concentration showed increasing effect on chlorophyll content (in fresh leaves), total soluble sugar and Ascorbic acid content in fruit, while 8.0 and 10.0 ppm show non-significant increase in the above parameters.

Key words- Tomato, miraculan, chlorophyll, total soluble sugar, ascorbic acid

Introduction

Bioregulators currently account for only a small share (5 to 10%) of total world wide agrochemical in market and they form only a small portion of total sales of agrochemicals world wide (Raghava and Raghava, 1990; 1991). In which, one of the recently used plant growth regulators is long chain aliphatic alcohol. TRIA is a C-30 primary alcohol was first identified in the early 1933 as a natural constituent of plant waxes (Kolkar, 1978) in alfa-alfa meal (Ries, *et al.* 1977) were first isolated in form of crystalline colourless substance. TRIA is available in market in various trade names, viz. Vipul, Miraculan, Mixtalol, Golden vipul, Paras, Tween 20, Jeevan and Nutron, etc. Out of these long chains aliphatic alcohols Mixtalol (a mixture of long chain aliphatic alcohols) are very common in use due to their growth regulating properties contents; leaf area, crop yield, biomass, controlling stomatal opening and decreasing photorespiration in many horticultural and other crops (Zelitch, 1975; Ries, *et al.* 1977; Ries and Wert, 1982; Nickell, 1982; Menon and Srivastava, 1984; Ries, *et al.* 1990; Raghava and Raghava, 1991; Malik *et al.*, 1993; Zhou and Zhao, 1995; Jiang and Sun, 2011). Since various investigations revealed that TRIA affect several metabolic processes in different plants as mentioned above i.e. ultimately influenced the various biochemical parameters which are reported by different workers on different plants (Debta and Murty, 1981; Houtz *et al.*, 1985b; Minzan and Xiuhua, 2009; Mishra and Raghav, 2010). The present study was carried out to analyse the influence of TRIA (Miraculan) on biochemical parameter in tomato.

Materials and Methods

The certified seed of tomato (*Lycopersicon esculantum* Mill.) were obtained from Agriculture Extension Division of IARI, Pusa campus, New Delhi. The seed were surface sterilized with mercuric chloride, washed thoroughly with distilled water and soaked in water for the duration of 24 hr. before sowing. There after, 5 kg well processed and sieved black soil and manure mixture were filled in pots and moist it

for sowing. Five healthy seeds were sown in each pot and after germination maintained one plant at 15th days after sowing. The pots were irrigated and germinated plants with pots were kept in light till the end of experimental trial, after 15 days of sowing leaves were well developed on the plant. The eight treatment combination as concentration (Control, 0.5, 1.0, 2.0, 3.0, 4.0, 6.0, 8.0 and 10.0 ppm) of Miraculan was applied as foliar application at 20, 40 and 60 days after sowing (DAS) (i.e. preflowering, flowering and post flowering stage) of tomato crop. The effect of Miraculan was observed at 25, 45, and 65 days after sowing for chlorophyll (ppm) content in leaves and total soluble sugar, ascorbic acid content in fruit at maturity stage. Chlorophyll was analysed in collected fresh leaf samples at each stage of method described by Arnon (1949) and Jensen & Jensen (1972) in fresh leaves. Total soluble sugar (mg/l) content was estimated in fresh fruit of tomato by the method of Morris (1948) and High Kin and Frenkel (1962), ascorbic acid (mg/l) content was analysed in fresh fruit by the method proposed by Dhopte and Manuel (1989). The collected data from biochemical analysis were subjected to statistical analyses of variance using completely randomised block design as described by Pense and Sukhatme (1995).

Results and Discussion

The leaves collected at three different growth stages i.e. 25, 45 and 65 DAS. Increased chlorophyll content (ppm fresh wt.) was found in all the concentrations significantly over the control at all stages but increasing levels of Miraculan from 0.5 ppm up to 2.0 ppm were enhanced the chlorophyll content of leaves in all the observation. The application of Miraculan at higher concentration from 4.0 ppm up to 10.0 ppm was showed significant decreasing the chlorophyll content in the leaves at all the growth stage of crop. The maximum chlorophyll content 13.26 ppm, 14.69 ppm and 14.99 ppm were significantly recorded with application of 2 ppm miraculan on fresh leaves at 25, 45 and 65 DAS respectively (Table - 1). Similarly the total soluble sugar and ascorbic acid content in fruits showed the increasing effect with same concentrations (table-2). At level of miraculan 2 ppm foliar application was significantly increased 4.78 mg/l of total soluble sugar content and 92.83 ppm of ascorbic acid content in fruit among all other miraculan doses at maturity stage of fruit (Table -2).

Table- 1 Effect of Miraculan on the chlorophyll (ppm) content in the leaves of Tomato at different stages of growth (all the datas are average of three replicate)

Conc./DAS	25	45	65
Control	10.45 ±0.68	11.05 ±0.99	11.65 ±0.98
0.5	11.48 ±0.77	11.94 ±0.76	12.16 ±0.96
1	11.98 ±0.76	12.56 ±0.75	12.85 ±0.83
2	13.26 ±0.65	14.69 ±0.76	14.99 ±0.54
4	11.95 ±0.68	12.42 ±0.45	12.55 ±0.96
6	10.96 ±0.77	11.25 ±0.73	11.96 ±0.78
8	10.25 ±0.69 *	10.95 ±0.85 *	11.01 ±0.76 *
10	9.85 ±1.43 *	10.62 ±1.06 *	10.50 ±1.21 *
C.D. at 5% Level	0.64	0.48	1.15

Table- 2 Effect of miraculan on the total soluble sugar (mg/l) and Ascorbic Acid (mg/l) Content in fruits of Tomatol at maturity (all the datas are average of three replicate)

Conc. (ppm)	Total Soluble Sugar	Ascorbic Acid
Control	2.98 ± 0.06	88.82 ± 0.63
0.5	3.32 ± 0.03	89.94 ± 0.34
1	3.99 ± 0.04	90.25 ± 0.74
2	4.78 ± 0.02	92.83 ± 0.38
4	3.36 ± 0.24	90.42 ± 0.34
6	3.30 ± 0.32	89.01 ± 0.82
8	2.86 ± 0.04*	88.60 ± 0.84*
10	2.84 ± 0.03*	87.80 ± 0.19*
C.D. at 5% level	0.35	1.47

- * Non-significant, DAS =Days After Sowing, Conc. = Concentration

The increase in chlorophyll content by the application of 2.0 ppm miraculan was due to inhibition of senescence or iron uptake which regulated chlorophyll biosynthesis (Menon and Shrivastava, 1984) and appeared dense green coloured leaf. TRIA universally improves the photosynthetic efficiency of plant (Chaudhary et al. 2006), increase leaf area and its contents of chlorophyll and protein in seeds as well as control stomatal opening (Yadava and Sreenath, 1975; Zelitch, 1975; 1979; Eriksen, 1981, Menon 1987; Setia *et al.*, 1989; Gupta *et al.*, 2000 and Xingping Chen *et al.*, 2002)). However, the general effect of TRIA appears to increase the dry weight of the plants (Satler and Thiman, 1980, Raghava and Raghava, 1991; Ries, 1991; Raghava *et al.* 2007). So, the results showed correlations with the work of different above workers the increases due to acceleration of metabolic biochemical complex mechanism involve which in promoted by TRIA in the plant.

Conclusion

The application of miraculan as a plant growth regulator to express the tomato growth was found significant increased chlorophyll content in leaves and total soluble sugar and ascorbic acid content in fruit but the 2.0 ppm concentration as foliar application was found suitable for vigorous and blooming growth of tomato. The other doses of miraculan i.e. 4.0, 6.0, 8.0 and 10.0 ppm could be needed for new scope of further research.

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References

1. Arnon, D.I. (1949). Copper enzyme in isolated chloroplasts: Polyphenol Oxidase in *Beta vulgaris*, *Plant Physiol.* 24: 1-5.

2. Chen, XinPing, Yuan, Hongyu, Chen; Rangzhi, Zhu Lili, He, G., Chen X. P., Yuan H. Y., Chen, R. Z., Zhu L.L. and He, G.C. (2003). Biochemical and photochemical changes in response to triacontanol in rice. *Plant Growth Regu.* 40(3):249-256.
3. Chibnall, A.C., Williams, A.L., Latner, A.L. and Piper, S.N. (1933). The isolation of n-triacontanol from Lucerne wax. *Biochemistry Journal.* 27: 183-188.
4. Choudhury, B. (1996). Solanaceous fruits. *Vegetables*, National Book Trust, India, 43-69.
5. Debata, A. and Murty, K.S. (1981). Effect of Growth Regulators on Photosynthetic efficiency, Translocation and Senescence in Rice. *Indian Journal of Experimental Biology.* 19: 986-987.
6. Dhopte, A.M. and Manuel, L.M. (1989) Useful Techniques for Plant Scientists. Publication of Forum for Plant Physiologists, Akola (Maharashtra), India.
7. Di Cui, Minzan Li and Xiuhua Li, (2009). IFIP advances in information and communication technology, 2009 Vol. 295, Computer and computing technologies in agriculture II, 3:2111-2118.
8. Eriksen, A.B., Sellden, G., Skogen, D. and Nitsens. (1981). Comparative analysis of the effect of triacontanol on photosynthesis, photorespiration and growth of tomato (C3 plant) and maize (C4 plant). *Planta.* 152: 44-49.
9. Gupta, N.K., Jain, S.K., Gupta, S. and Sen, N.L. (2000). Plant Bioregulators: Potential tool to enhance fruit production in India. In K.K. Bora, K. Singh and A. Kumar edited, *Seeds, Bioregulators and Applied Plant Biotechnology*. Pointer Publishers, Jaipur, India, pp. 1 - 22.
10. Highkin, H.R. and Frenkel, A.W. (1962) Studies of growth and metabolism of a barley mutant lacking Chl-b. *Plant Physiol.*, 37 (6):814-820.
11. Houzt, R.L., Ries, S.K. and Tolbert, N.E. (1985). Effect of triacontanol on Chlamydomonas. II. Specific activity of ribulose-biphosphate carboxylase/oxygenase, ribulosebi-phosphate concentration and characteristic of photorespirations. *Plant Physiol.* 79: 365-370.
12. Jenson, S.L. and Jenson, A. (1972). Quantitative determination of carotenoids in photosynthetic tissue. In San Pietro (ed.), *Methods in Enzymology.* 23 : 218-228.
13. Malik, C.P., Setia, R.C. and Setia, N. (1993). Role of Plant Growth Regulators in Increasing Crop Productivity and the Indian Scenario, In S.S. Purohit edited. *Hormonal Regulation of Plant Growth and Development.* 6 : 161 - 197.
14. Menon K.K.G. (1987). Harnessing nature towards better agriculture. In Srivastava, H.C., Bhaskaran, S. and Menon, K.K.G., edited, *Crop productivity*, Oxford & IBH Publishing Company Private Limited, New Delhi , pp. 19-33.
15. Menon, K.K.G. and Srivastava, H. (1984). Increasing plant productivity through improved photosynthesis *Proceedings of Indian Academy of sciences (Plant Sci.)* . 93: 359 - 378.
16. Morris, D. L. (1948) Quantitative determination of carbohydrates with Dreywood's anthrone reagent. *Science*, 107 254-255.
17. Nickell LG (1982) Plant Growth Regulators- Agricultural uses. Springer- Verlag New York.
18. Panse, V.G. and Sukhatme, P.V. (1985). *Statistical methods for agricultural workers* (4th ed.) ICAR, New Delhi.
19. Raghava, R. P, B. P. Singh, N. Raghava, S. P. Singh and B. P. Gupta (2007) Physiological approaches for higher yield cape gooseberry. *Indian j. Applied & Pure Bio.* 22 (1) : 27-30.
20. Raghava, R.P. and Raghava, N. (1990). Use of bioregulators in orchards and nurseries. In, Ram Parkash edited, *Advances in Forestry Research In India*. International Book Distributors (India), Dehradun, 6: 19-54.
21. Raghava, R.P. and Raghava, N. (1991). Use of Mixtalol and Triacontanol as bioregulators, *Everyman's Sciencei.* 26 : 51-52.

22. Ries, S.K. (1985). Regulation of plant growth with triacontanol. *CRC Critical Review in Plant Science*. 2: 239-285.
23. Ries, S.K. and Wert, V. (1982). Rapid effects of triacontanol *in vivo* and *in vitro*. *Journal of Plant Growth Regulator*. 1: 117-127.
24. Ries, S.K., Wert, B., Sweeley, C.C. and Leavitt, R.A. (1977). Triacontanol : A new naturally occurring plant growth regulators. *Science*. 195: 1339-1341.
25. Ries, S.K., Wert, V.F., O'Leray, N.F.D. and Nair, M. (1990). 9- β -L(+) adenosine: A new naturally occurring plant growth substance elicited by triacontanol in rice. *Plant Growth Reg.* 9(3) : 263-273.
26. Setia, D.C., Richa, Setia, N., Ahuja, K.L. and Malik, C.P. (1989). Effect of mixtanol on growth and yield components of Indian mustard (*Brassica juncea*). *Plant Growth Regulation*. 8 (2): 185-192.
27. W.J. Jiang, M. Sun (2011). Guang Pu Xue Yu Guang Pu Fen Xi 2011 Mar; 31 (3) 758-761 Yadava R.B.R. and Sreenath, P.R. (1975). Influence of some growth regulators. *Indian J. Plant Physiol.*, 19, 135-139.
28. Yadava, R.B.R. and Sreenath, P.R. (1975). Influence of some growth regulators. *Indian J. Plant Physiol.*, 19, 135-139.
29. Zelitch, I. (1975). Improving the efficiency of Photosynthesis. *Science*. 188: 205.
30. Zhou, W. Tao, S and Zhao, D. (1995). Physiological Regulation of Mixtanol in Rape Senescence and its yield effects. *Plant Growth Regul.* 14:37-40.

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